AMENDMENTS TO THE CLAIMS

1. (Currently amended) A method for the construction of randomized gene libraries in ~~suitable~~ yeast cells sensitive to *Kluyveromyces lactis* γ toxin and which are capable of homologous recombination comprising the following steps:

introducing into said yeast cells ~~capable of homologous recombination~~;

a) a target vector comprising a first DNA sequence coding for at least a γ-subunit of a *Kluyveromyces lactis* killer toxin as negative selection marker, said DNA sequence being flanked at its 5' end by a first target sequence and at its 3' end by a second target sequence and;

b) a donor DNA sequence which is flanked at its 5' end by a DNA sequence which is homologous to said first target sequence and flanked at its 3' end by a DNA sequence which is homologous to said second target sequence;
and

cultivation of said yeast cells under ~~suitable~~ conditions allowing the selection of cells in which said DNA sequence in the target vector encoding at least a γ -subunit of a *Kluyveromyces lactis* killer toxin has been replaced by said donor sequence by means of homologous recombination thereby abolishing expression of said γ -subunit of a *K. lactis* killer toxin.

2. (Previously presented) The method of claim 1, wherein said target vector further comprises a second DNA sequence encoding at least one protein region.

3. (Original) The method of claim 2 wherein said first DNA sequence of said target vector encoding at least the γ -subunit of the *K. latics* killer toxin and being flanked by said two target sequences replaces a protein region encoding DNA sequence of said second DNA sequence comprised in said target vector.

4. (Previously presented) The method of claim 1 wherein said DNA sequence encoding at least the γ subunit of the *K. lactis* killer toxin is under control of a heterologous promoter.

5. (Original) The method of claim 4 wherein said promoter is located between the DNA sequence encoding at least the γ subunit of *K. lactis* killer toxin and one of the two target sequences.

6. (Currently amended) The method of claim 1, wherein said first DNA sequence of said target vector comprises at least one restriction enzyme ~~unique~~ recognition site that is unique for a given restriction enzyme.

7. (Previously presented) The method of claim 6, wherein said unique recognition site is located in the coding region of the γ-toxin DNA sequence.

8. (Currently amended) The method of claim [[1]] 2 wherein said ~~first~~ second DNA sequence encodes an antibody or a single chain antibody.

9. (Previously presented) The method of claim 8 wherein said first DNA sequence of said target vector replaces at least one CDR region of said antibody or said single chain antibody.

10. (Original) The method of claims 8 or 9 wherein said first DNA sequence comprising the γ subunit of *K. lactis* killer toxin is transcribed in the opposite direction of said antibody or single chain antibody gene.

11. (Previously presented) The method of claim 1 wherein said γ-toxin subunit of the *K. lactis* killer toxin lacks a signal peptide.

12. (Canceled)

13. (Previously presented) The method of claim 1 wherein said target vector is introduced into said host cells in linearized form.

14. (Currently amended) The method of claim 13 wherein said target vector is linerarized by cutting with a restriction enzyme recognizing in said first DNA sequence of said target vector said at least one ~~unique~~ restriction enzyme recognition site that is unique for a given restriction enzyme.

15. (Previously presented) The method of claim 1 wherein said donor sequence comprises a DNA sequence encoding a protein region.

16. (Previously presented) The method of claim 1 wherein said target vector and said donor sequence are introduced into said host cells by co-transformation.

17. (Previously presented) The method of claim 12 wherein said yeast cells are cultivated at a temperature selected from the range of 24°C to 30°C.

18. (Canceled)

19. (Withdrawn) Use of a *Kluyveromyces lactis* killer toxin γ-subunit as negative selection marker for the construction of randomized gene libraries and/or region replacement by homologous recombination.

20. (Withdrawn) A DNA vector which comprises the following sequences: a first target sequence for homologous recombination, a *TEF* promoter from *Ashbya gossypii* driving transcription of a *K. lactis* killer toxin, a DNA sequence encoding at least a γ-subunit of a *K. lactis* killer toxin and a second target sequence for homologous recombination.

21. (Withdrawn) A host cell comprising a vector of claim 20.

22. (Previously presented) The method of claim 4 wherein the promoter is a constitutive promoter.

23. (Previously presented) The method of claim 4 wherein the promoter is a TEF promoter from *Ashbya gossypii*.

24. (Currently amended) The method of claim 6 wherein the ~~unique~~ restriction enzyme recognition site that is unique for a given restriction enzyme is located between the coding region of the γ-toxin DNA sequence and the promoter.

25. (Previously presented) The method of claim 9 wherein the first DNA sequence of said target vector replaces a CDR3V_L region of said antibody or said single chain antibody.

26. (Previously presented) The method of claim 9 where the first DNA sequence of said target vector replaces a CDR2 and a CDR3 region at said antibody or said single chain antibody.

27. (Previously presented) The method of claim 12 wherein said host cells are *Saccharomyces cerivisiae* cells.

28. (Previously presented) The method of claim 15 wherein said donor sequence comprises a DNA sequence encoding a CDR region of an antibody.

29. (Withdrawn) The host cell of claim 20 which is a yeast cell.

30. (Withdrawn) The host cell of claim 29 which is a *Saccharomyces cerevisiae*.

31. (Previously presented) The method of claim 2, wherein said second DNA sequence encodes more than two protein regions.

32. (Previously presented) The method of claim 2, wherein said second DNA sequence encodes a full length protein.

33. (Previously presented) The method of claim 15, wherein said protein region is a CDR region of an antibody.